

Evaluation of Melatonin Hormone and Nitric oxide Levels in Non-alcoholic Fatty Liver Patients in Relation to Obesity and Oxidative Stress

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Abstract:

Objective: To assess the levels of melatonin hormone and oxidative stress markers(TBARS, MDA and GSH) in NAFLD patients and obese subjects and compared with healthy group.

Material and Methods: A total of (63) volunteers inserted in study, divided to three groups :group A :(21) obese patients with NAFLD, group B: (21) obese subjects and group C: (21) healthy subjects. Melatonin hormone, TBARS, MDA and GSH were measured.

Results: The current study recorded that there was a significant decrease in melatonin hormone in both group A & B when compare to control ($p > 0.05$). The mean of GSH also significantly decrease in NAFLD and obese patients when compared to healthy subject ($p > 0.05$). Both TBARS and NO were significantly rise in group A among to other groups($p > 0.05$), while MDA not show and difference between studied groups

Conclusion:Melatonin hormone effected by both NAFLD and obesity.

Key words: *Melatonin, TBARS, MDA, GSH, obesity*

INTRODUCTION:

Non-alcoholic fatty liver disease (NAFLD) is one of the most famous causes of chronic liver impairment. It is distinguished by asymptomatic increase in serum enzymatic liver levels but not used excessive alcohol intake or suffered from other chronic liver diseases. [1]. NAFLD is most common in obese patients with type 2 diabetes mellitus and insulin resistance [2]. Recent estimations suggest that about 20%–25% of the populations are obese. The percentage of NAFLD in the obese subjects can be about 95% depending on the degree of the obesity. As the prevalence of obesity increases, the prevalence of NAFLD will increase, making it the most common cause of chronic liver disease [3].

NAFLD encompasses a range of clinic-pathologic exist, all of which involve fat accumulation in the liver parenchyma. Nonalcoholic steatohepatitis (NASH) is a subtype of NAFLD association with hepatocyte necrosis with or without fibrosis; it develop to chronic liver disease and cirrhosis [4]. In NAFLD patients appear that fat diffusion inside liver tissue and secreted hepatic steatosis and take place in the absence of alcohol intake. NAFLD is a serious cause of concern because of its direct association with diabetes, obesity and hyperlipidemia [5]. Simple steatosis is shown around the liver and then leads to the proliferation of fat cells into hepatocytes causing nonalcoholic steatohepatitis (NASH), which develops into cirrhosis of the liver [6]. Obesity is a major risk factor lead to the development of NAFLD [7]. Fat-rich food is the (first hit) causing NAFLD and inflammation is the (second hit) leading to convert NAFLD to NASH [8]. Currently, there are no direct treatments for liver in the case of NAFLD but all therapies targeting to reduce the risk of NAFLD [9].

Melatonin is a hormone produced by the pineal gland. The secretion of melatonin is controlled by light intensity [10]. Melatonin play an important role in multiple biological functions and acts as an anti-oxidant [11]. It has act as anti-inflammatory molecules [12], regulates heart rate [13] and support the human immunity [14]. It also has anti-cancer properties [13]. Recent

study suggested that melatonin also regulated the lipid metabolism [16], decrease insulin resistance [17], control glucose levels [18], and decrease body weight [19]. This study was designed to evaluate the effect of NAFLD on melatonin levels and oxidative stress markers.

MATERIAL AND METHODS:

The study was conducted at the Center of Liver and Gastrointestinal Diseases Baghdad Teaching Hospital. The participants included 63 volunteers divided into three groups: the first group (21) volunteers, obese patients (BMI > 30) with NAFLD, and the second group (21), obese volunteers (BMI > 30) without other disease. Third (21) volunteers are control group. Blood samples were collected from fasting patients in the morning (8:00 am - 10:00 am). The time of sample taken was affected on our results. A total of (5 ml) was draw from venous and transfer into the tubes and left for (15 min.) and then placed in the centrifuge for (10 min) and then separated the serum and stored in frozen degree (-20) until the day of the analysis.

Measurements:

Serum melatonin hormone and oxidative stress marker (glutathione (GSH), Thiobarbituric acid reactive substances (TBARS), Malondialdehyde (MDA)) and nitric oxide (NO) were measured.

Melatonin was measured quantitatively by using ELISA kit (CUSABIO, China) . GSH levels were measured according to Julins (1994) method (20). TBARS and MDA levels were determined spectrophotometrically according to Alam Zeb (2016) methods (21).

RESULTS:

The current study shows that there were a significant difference in both ALT and AST in group A when compared to other studied groups. Alkaline phosphatase show no difference among groups.

Table (1)

	No	ALT	AST	ALP
Group A	21	47.14±22.52 a	49.65±15.39a	95.1±22.41 a
Group B	21	28.61±7.90 b	32.54±8.66 b	96.19±24.64a
Group C	21	27.80±6.35 b	34.32±7.41b	96.71±25.29a
P value	63	0.002	0.004	0.093
LSD		3.4136	11.226	0.447

Table (2)

	No	Melatonin	GSH	TBARS	MDA	NO
Group A	21	12.24±5.45b	65.75±17.59a	7.61±0.58a	5.53±0.94 a	122.76±59.78a
Group B	21	12.61±4.90b	73.34±18.53b	6.90±0.64b	5.99±1.04a	95.42±38.67b
Group C	21	17.80±6.15a	80.62±18.41c	5.51±0.90c	5.91±0.86a	79.76±21.57b
P value	63	0.002	0.04	0.003	0.25	0.001
LSD		3.4136	11.226	0.447	0.5901	26.541

The current study recorded that there was a significant decrease in melatonin hormone in both group A & B when compare to control ($p > 0.05$). The mean of GSH also significantly decrease in NAFLD and obese patients when compared to healthy subject ($p > 0.05$). Both TBARS and NO were significantly rise in group A among to other groups($p > 0.05$), while MDA not show and difference between studied groups, table (2).

DISCUSSION:

Several studies have evaluated that melatonin treatment decreased body weight in animal and human [22,23]. A recent study recorded that melatonin has action on inflammatory process, fat degradation, and bioenergetic reaction in animals[24]. Current study found that melatonin significantly decreased body weight in patients when compared to control group. Melatonin act as controlling factor in glucose metabolism. Two types of melatonin receptors called MT1 and MT2, which called G-protein-coupled receptors[25]. Each receptors are located in the islet of Langerhans and are participated in the controlling of glucagon production from α -cells and insulin releasing from β -cells [25]. Other study suggested that stimulating of MT1 or MT2 act as an inhibitor for second messenger cAMP or cGMP accompanied by decrease insulin production [26, 27].

This study had many limitations. Firstly, it is not recorded that the food intake for volunteers, which was one of the limitations of the study.

Decreased in body weight could lead to a **decline** of adiposity and fat accumulation, which lead to decrease in hepatic steatosis. A decrement of steatosis, in turn, down-controlled inflammatory markers. Recent study suggest that melatonin treatment was cause weight reduction, which could be elucidated the melatonin action on body mass, or other **advantageous** actions. However, melatonin as therapeutic agent in NAFLD patients as a result of both anti-inflammation and anti-obesity effects[28].

Oxidative stress defined as inequality between defense factors which include superoxide dismutase (SOD) reduced glutathione (GSH), and reactive oxygen species

(ROS) include superoxide. Reduction of antioxidants molecules as glutathione, β -carotene, or vitamin C and E or rise increasing of ROS in the liver may occur in nonalcoholic steatohepatitis (NASH). The contribution of serum stress parameters in liver cirrhosis is unclear for hepatocellular carcinoma. Oxidative stress markers (serum levels of 8-hydroxy-2-deoxyguanosine 8-OHdG or 4-hydroxy-2-nonenal 4HNE), have been recorded to be elevated and antioxidants (superoxide dismutase) decline in NASH patients; however, no significant difference in serum 8-OHdG levels between NASH and FL was reported [29,30]

Nitric oxide (NO), which has short half-life free radical elaborate in different biological functions, consist of vascular balance, neurotransmission and host protection, is result from L-arginine by the action of nitric oxide synthase (NOS) enzyme. There are three isoforms of NOS (neuronal, endothelial, and inducible NOS) have been known. However, when there is increase in the NO production which lead to induce the inflammation-associated tissue damage, which reported a major role in an inducible NOS-mediated NO secretion. Garcia-Monzon and colleges suggest that

the pathogenesis of NASH, NO has a cytotoxicity effect by effected on superoxide anions to produce peroxynitrite, a strong oxidant that activate nitration of tyrosine to produced nitrotyrosine[31]. The NOS was found to be significantly elevated in the advance form of NASH; NO has been suggested to have a major role in the process of fibrosis in NASH patient[32].

CONCLUSION:

This study concluded that melatonin level decrease in NAFLD patient and obese subject when compared to healthy subjects. All oxidative stress markers effected by both obesity and NAFLD disease except MDA.

REFERENCES:

- Teli MR, James OF, Burt AD, Bennett MK, Day CP. The natural history of nonalcoholic fatty liver: a follow-up study. *Hepatology* 1995;22:1714–1719.
- Haider Qasim Hamood. Radiation Physics and its applications in diagnostic radiological techniques. 1st edition, December 2017.
- Dixon JB, Bhathal PS, O'Brien PE. Nonalcoholic fatty liver disease: predictors of nonalcoholic steatohepatitis and liver fibrosis in the severely obese. *Gastroenterology* 2001;121:91–100
- Powell EE, Cooksley WG, Hanson R, Searle J, Halliday JW, Powell LW. The natural history of nonalcoholic steatohepatitis: a follow-up study of forty-two patients for up to 21 years. *Hepatology* 1990;11:74–80.
- Rinella ME. Nonalcoholic fatty liver disease: a systematic review. *JAMA*. 2015;313(22):2263–2273.
- Yeh MM, Brunt EM. Pathology of nonalcoholic fatty liver disease. *Am J Clin Pathol*. 2007;128(5):837–847.
- Cohen JC, Horton JD, Hobbs HH. Human fatty liver disease: old questions and new insights. *Science*. 2011;332(6037):1519–1523.
- Day CP, James OF. Steatohepatitis: a tale of two "hits"? *Gastroenterology*. 1998;114(4):842–845.
- Marchesini G, Petta S, Dalle Grave R. Diet, weight loss, and liver health in nonalcoholic fatty liver disease: Pathophysiology, evidence, and practice. 2015.
- Skene DJ, Arendt J. Human circadian rhythms: physiological and therapeutic relevance of light and melatonin. *Ann Clin Biochem*. 2006;43(Pt 5):344–353.
- Manchester LC, Coto-Montes A, Boga JA, Andersen LP, Zhou Z, Galano A, Vriend J, Tan DX, Reiter RJ. Melatonin: an ancient molecule that makes oxygen metabolically tolerable. *J Pineal Res*. 2015;59(4):403–419.
- Borges Lda S, Dermargos A, da Silva Junior EP, Weimann E, Lambertucci RH, Hatanaka E. Melatonin decreases muscular oxidative stress and inflammation induced by strenuous exercise and stimulates growth factor synthesis. *J Pineal Res*. 2015;58(2):166–172.
- Lee JS, Cua DJ. Melatonin Lulling Th17 cells to sleep. *Cell*. 2015;162(6):1212–1214.
- Di Bella G, Mascia F, Gualano L, Di Bella L. Melatonin anticancer effects: review. *Int J Mol Sci*. 2013;14(2):2410–2430.
- Sun H, Huang FF, Qu S. Melatonin: a potential intervention for hepatic steatosis. *Lipids Health Dis*. 2015;14:75.
- Peschke E, Bahr I, Muhlbauer E. Melatonin and pancreatic islets: interrelationships between melatonin, insulin and glucagon. *Int J Mol Sci*. 2013;14(4):6981–7015
- Espino J, Pariente JA, Rodriguez AB. Role of melatonin on diabetes-related metabolic disorders. *World J Diabetes*. 2011;2(6):82–91.
- Eckel RH, Depner CM, Perreault L, Markwald RR, Smith MR, McHill AW, Higgins J, Melanson EL, Wright KP, Jr. Morning circadian misalignment during short sleep duration impacts insulin sensitivity. *Curr Biol*. 2015;25(22):3004–3010.
- Julius M, Lang CA, Gleiberman L, Harburg E, DiFrancesco W, Schork A. *Glutathione and morbidity in a community-based sample of elderly. J Clin Epidemiol* 1994;47:1021–6.
- Alam Zeb and Fareed Ullah. A Simple Spectrophotometric Method for the Determination of Thiobarbituric Acid Reactive Substances in Fried Fast Foods. *Journal of Analytical Methods in Chemistry*. Volume 2016 (2016), Article ID 9412767, 5 pages
- Chojnacki C, Walecka-Kapica E, Klupinska G, Pawlowicz M, Blonska A, Chojnacki J. Effects of fluoxetine and melatonin on mood, sleep quality and

- body mass index in postmenopausal women. *J Physiol Pharmacol.* 2015;66(5):665–671.
22. Favero G, Stacchiotti A, Castrezzati S, Bonomini F, Albanese M, Rezzani R, Rodella LF. Melatonin reduces obesity and restores adipokine patterns and metabolism in obese (ob/ob) mice. *Nutr Res.* 2015;35(10):891–900
 23. Agil A, Rosado I, Ruiz R, Figueroa A, Zen N, Fernandez-Vazquez G. Melatonin improves glucose homeostasis in young Zucker diabetic fatty rats. *J Pineal Res.* 2012;52(2):203–210.
 24. Catta-Preta M, Mendonca LS, Fraulob-Aquino J, Aguila MB, Mandarim-de-Lacerda CA. A critical analysis of three quantitative methods of assessment of hepatic steatosis in liver biopsies. *Virchows Arch.* 2011;459(5):477–485.
 25. Legros C, Devavry S, Caignard S, Tessier C, Delagrangre P, Ouvry C, Boutin JA, Nosjean O. Melatonin MT(1) and MT(2) receptors display different molecular pharmacologies only in the G-protein coupled state. *Br J Pharmacol.* 2014;171(1):186–201.
 26. Kemp DM, Ubeda M, Habener JF. Identification and functional characterization of melatonin Mel 1a receptors in pancreatic beta cells: potential role in incretin-mediated cell function by sensitization of cAMP signaling. *Mol Cell Endocrinol.* 2002;191(2):157–166.
 27. Sartori C, Dessen P, Mathieu C, Monney A, Bloch J, Nicod P, Scherrer U, Duplain H. Melatonin improves glucose homeostasis and endothelial vascular function in high-fat diet-fed insulin-resistant mice. *Endocrinology.* 2009;150(12):5311–5317.
 28. Huda JaberWaheed ,MostafaAbdulfatahShafek and Huda Abdul RidhaHadi. An Essential Role for TGF- β 1 and other Clinical Parameters in Non-alcoholic Fatty Liver Disease in Iraqi Patients. *International Journal of Current Microbiology and Applied Sciences* ISSN: 2319-7706 Volume 5 Number 9 (2016) pp. 650-658.
 29. Hang Sun, Xingchun Wang, Jiaqi Chen,2 Kexiu Song,1 Aaron M. Gusdon,3 Liang Li,1 Le Bu,1 and Shen Qu. Melatonin improves non-alcoholic fatty liver disease via MAPK-JNK/P38 signaling in high-fat-diet-induced obese mice. *Lipids Health Dis.* 2016; 15: 202.
 30. Narasimhan S, Gokulakrishnan K, Sampathkumar R, Farooq S, Ravikumar R, Mohan V, Balasubramanyam M. Oxidative stress is independently associated with non-alcoholic fatty liver disease (NAFLD) in subjects with and without type 2 diabetes. *ClinBiochem.* 2010 Jul;43(10-11):815-21.
 31. Garcia-Monzon C, Majano PL, Zubia I, Sanz P, Apolinario A, Moreno-Otero R. Intrahepatic accumulation of nitrotyrosine in chronic viral hepatitis associates with histological severity of liver disease. *J Hepatol* 2000; 32: 331–8
 32. Yoneda M, Hotta K, Nozaki Y, et al. Influence of inducible nitric oxide synthase polymorphisms in Japanese patients with non-alcoholic fatty liver disease. *Hepatol Res* 2009; 39: 963–71.