Therapeutic Effect of Sildenafil in Experimental Colitis through Anti-Oxidative Stress and Inhibition of Adhesion Molecules

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Abstract
Inflammatory bowel disease is a progressive inflammatory intestinal disease characterized by colon tissue edema and increased epithelial permeability of colon and extensive infiltration of leukocytes in the colon. Failure of current treatment strategies and the adverse effects of currently used drugs, it is crucial to investigate new drugs with more favourable therapeutic profiles. The aim of the present study was to evaluate possible therapeutic effects of sildenafil against acetic acid induced colitis in a rat model and to found out correlation between severity index with oxidative stress parameters and inflammatory markers. Experimental colitis was induced in rats by rectal administration of 4% acetic acid (vol/vol). Rats with colitis were received either sildenafil 10mg/kg or sulfasalazine 100mg/kg orally for 7 days. Macrosopic and microscopical assessment and the measurement of the colonic cytokines (IL-6 and TNF-α), oxidative stress markers; myeloperoxidase (MPO) and malondialdehyde (MDA), and adhesion molecules (E-Selectin and ICAM-1). Our findings had shown that both macroscopical lesion area and histological colonic injury induced by acetic acid were significantly reduced by both sildenafil and sulfasalazine. These were accompanied by attenuation of the elevated colonic MPO activity, MDA and proinflammatory cytokines. Besides downregulation of the adhesion molecules. These results demonstrated that sildenafil possesses therapeutic potential in experimental colitis. The anti-inflammatory actions involve antioxidant effect along with inhibition of adhesion molecule synthesis in the colonic tissues.

Key Word: Sildenafil, ulcerative colitis, Oxidative stress, Adhesion molecules

INTRODUCTION
Inflammatory bowel disease (IBD) is a chronic inflammatory disease. It is of two different types; ulcerative colitis and Crohn’s disease [1]. They are characterized by immune-mediated chronic relapsing intestinal disorders, associated with rectal bleeding and diarrhea resulting in damage of the epithelial barrier and the formation of epithelial ulceration[2]. Despite the fact that little is known about etiology of these diseases, it is believed to involve an abnormal host response to endogenous or environmental or immunological factors, causing initial tissue damage followed by amplification of this response [3]. IBD are also associated with extensive inflammatory infiltrates in the lamina propria characterized by extensive inflammatory infiltrates consisting of polymorphonuclear neutrophils, eosinophils and plasma cells, leading to a remarkable production of unstable chemical species such as nitrogen and reactive oxygen species, significantly involved in injury[4,5]. Activation of these infiltrating cells results in the release of different pro-inflammatory mediators such as tumor necrosis factor-alpha (TNF-α), interleukin-1 β (IL-1 β), an IL-6, plays a crucial role in tissue disruption and ulceration [6]. Although extensive development has been made in the treatment of ulcerative colitis, adverse effects and incomplete efficacy of currently used medications is a continuous challenge[7]. For this reason, there is a need to develop new strategies that in could restore the altered immune response that emerges in the inflamed intestine.

One of the important strategies is targeting adhesion molecules that expressed on the surface of endothelial cells and lymphocyte; which in fact recruits immune and inflammatory cells from the periphery into site of inflammation. On the other hand there is evidence that nitric oxide (NO) inhibits the expression of adhesion molecules on endothelial cells, which is an important step in neutrophil migration[8]. Sildenafil is one of the drugs that have nitric oxide modulating activity[9]. This effect of sildenafil is attributable to inhibition of phosphodiesterase-type 5 enzyme that catalyze the hydrolysis of cGMP[10], and has vascular smooth muscle relaxant effect and inhibition of platelets aggregation by preventing the breakdown of nitric oxide –dependant cGMP[11]. Moreover sildenafil has antioxidative stress potential[12] and prevent lipid peroxidation and cytokine release[13].

MATERIAL AND METHODS
Materials
Animals: Adult male albino rats (200-220g) were purchased from animal house of the national center for drug control and researches (NCDCR). Animal were housed five per cage for one week prior to the experiment and had access to laboratory chow pellet and were allowed to drink tap water ad libitum. All animal experiments were performed after getting prior approval from the institutional animal ethics committee college of medicine Al-Nahrain university.
Drugs: sildenafil and sulfasalazine were purchased from Sigma–Aldrich company.

**Experimental Design**
This study was conducted on 40 adult male albino–wister rats weighing 200–220g previously submitted to starvation for at least 24hrs. Animals were divided into four group (n=10/group). Group I kept as control and received no treatment. Group II, III, IV were subjected to the induction of colitis by rectal administration of 4% acetic acid (AA) (v/v). Thirty minute after the induction of colitis group II was given normal saline orally; group III and IV were treated orally with sildenafil 10mg/kg and sulfasalazine 100mg/kg respectively for 7 days.

**Induction of colonic inflammation**
Since prior experiment has been shown to prevent the ulcerogenic action of certain drugs and chemical[14]. Rats were starved for at least 24hrs before the induction of colitis but were be allowed free access to tap water, during starvation, rats were kept in cages provided with a wide wire–mesh floor to avoid coprophagy. On the day of the experiment, water was held two hours before the procedure. Experimental ulceration in colon tissue was done according to the method described by Mousavizadeh et al[15] with slight modification. In brief, under light ether anesthesia rats were administered 5ml/kg of 4% acetic acid (AA) solution (BDH Chemical Ltd., England) by transrectally using a flexible silicone plastic tube with an external diameter of 2mm was inserted rectally into the colon to 8cm. After acetic acid administration, rats were holed horizontally for 2 min to prevent AA leakage. Control animals underwent the same procedure using equal volume of normal saline instead of AA solution.

**Preparation of drugs**
All drugs were freshly prepared before administration on the day of the experiment.
Investigated drug (Sildenafil) and the standard sulfasalazine were prepared as suspensions in distilled water using sodium Carboxymethyl cellulose (s CMC) 0.3% W/V. Sildenafil was used at a dose 10mg/kg (these doses were chosen depending on previous studies that have been showing their gastroprotective activity at a concentration 10mg/ml[16, 17]. Sulfasalazine was used as standard therapy in a dose of 100mg/kg[18].

**Assessment of colitis**
After the end of experiment, animals were sacrificed by an over dose of diethyl ether inhalation and then the abdomen was rapidly dissected and open and the colon was removed. The pieces of colons were cut open in an ice bath cleansed gently using normal saline, and observed normally for macroscopic and microscopic assessment. Then samples were cut into two pieces, one piece for histopathologic assessment (maintained in neutral formalin 10% as a fixater) and one piece for immunohistochemistry study.

**Macroscopic evaluation**
**Colonic mucosal damage** (mean area of colonic mucosal damage)
The excised colonic segment (8 cm proximal to anus) was immediately immersed in normal saline, cleaned from adherent tissues and then opened longitudinal and rinsed with 0.9% sodium chloride solution to discard the fecal materials. Then the segment was fixed with pins on a dissecting board, and the area of mucosal damage was measured using a computerized planimeter in accordance to the method described earlier[19].

**Colon edema** The colon specimen of each animal was incised along its mesenteric border and gently washed. This is measured through colon weight (CW). It was used as a index of tissue edema, which reflected the severity of colitis[20].

**Disease activity index (DAI)**
To quantify the clinical evaluation of the disease we used the DAI described by Meerveld and Tyler[21] that based which include body weight loss stool consistency, rectal bleeding (gross or occult) we used five grades of weight loss {0, no loss or weight gain; 1, 1-5 % loss; 2, 6-10%loss; 3 : 11-15% loss; 4: greater than 15% loss }, three grades of stool consistency {0: Normal; 2 : loose; 4 : diarrhea }, and three grades of bleeding {0 : normal; 2 occult blood –positive; 4 : gross bleeding }. The presence of occult blood in faces was determine using benzidine test. The total score of DAI was calculated as combined of these scores divided by 3[22].

**Macroscopic colonic score**
The macroscopic colonic score was assessed by the scoring system adapted from[23] as following: score are assigned based on the clinical features of the colon using a scale ranging from 0-4 as follows: 1, intact epithelium with no damage; 2, patch type superficial hyperemia; 3, generalized patch type hyperemic regions; 4, generalized hyperemic and hemorrhage.

**Histological evaluations:**
The colonic samples were fixed in 10% formalin, dehydrated, embedded in paraffin, deparaffinized with xylene, cut into 4 µm sections and stained by Hematoxylin and eosin (H&E). Slides were examined and scored for histopathological evaluation. The slides were coded to prevent observer bias during evaluation. All tissue sections were examined in a blinded fashion by experienced histopathologist and results scored according to Cooper et al[24].
### Table 1: Histopathological score of colitis

<table>
<thead>
<tr>
<th>Score</th>
<th>Destruction of epithelium and/or glandular crypts</th>
<th>Dilation of glandular crypts</th>
<th>Depletion and loss of goblet cells</th>
<th>Inflammatory cell infiltration</th>
<th>Edema</th>
<th>Hemorrhagic mucosa</th>
<th>Crypt abscess</th>
<th>Apoptosis</th>
<th>Dysplasia</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>morphologically normal</td>
<td>Normal aspect</td>
<td>Normal aspect</td>
<td>absence of infiltration</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
<td>absent</td>
<td>absent</td>
</tr>
<tr>
<td>1</td>
<td>focal destruction</td>
<td>Focal dilation</td>
<td>slightly depleted goblet cells</td>
<td>infiltrate at the sub-epithelial and lamina propria level or crypt bases infiltration</td>
<td>Focal</td>
<td>Focal</td>
<td>focal</td>
<td>Focal</td>
<td>Focal</td>
</tr>
<tr>
<td>2</td>
<td>zonal destruction</td>
<td>Zonal dilation</td>
<td>zonal or moderately depleted goblet cells</td>
<td>infiltration reaching muscularis mucosa</td>
<td>zonal</td>
<td>zonal and/or moderately diffuse</td>
<td>zonal</td>
<td>zonal</td>
<td>zonal</td>
</tr>
<tr>
<td>3</td>
<td>diffuse and/or mucosal ulceration involving submucosa and/or diffuse crypt loss</td>
<td>diffusely or complete depletion of goblet cells</td>
<td>severe and extensive infiltration reaching submucosa and/or involving muscularis propria</td>
<td>extensive and severe</td>
<td>Diffuse</td>
<td>Diffuse</td>
<td>Diffuse</td>
<td>diffuse</td>
<td>diffuse</td>
</tr>
</tbody>
</table>

### Table 2: Gross features in control and study groups

<table>
<thead>
<tr>
<th>Variable</th>
<th>Control n=10</th>
<th>Colitis n=10</th>
<th>Sild n=10</th>
<th>Sulfaz n=10</th>
</tr>
</thead>
<tbody>
<tr>
<td>MD(mm2)</td>
<td>0.00 ±0.00</td>
<td>16.11 ±0.74</td>
<td>7.65 ±0.44</td>
<td>5.26 ±0.43</td>
</tr>
<tr>
<td>CW (g)</td>
<td>1.13 ±0.17</td>
<td>3.12 ±0.24</td>
<td>2.81 ±0.18</td>
<td>1.63 ±0.23</td>
</tr>
<tr>
<td>DAI</td>
<td>0.00 ±0.00</td>
<td>10.50 ±1.50</td>
<td>4.40 ±1.11</td>
<td>2.10 ±0.30</td>
</tr>
<tr>
<td>MAC score</td>
<td>0.00 ±0.00</td>
<td>9.30 ±0.64</td>
<td>4.40 ±0.49</td>
<td>2.50 ±0.50</td>
</tr>
</tbody>
</table>

### Table 3: Histopathological score and adhesion molecules

<table>
<thead>
<tr>
<th>Variable</th>
<th>Control n=10</th>
<th>Colitis n=10</th>
<th>Sild n=10</th>
<th>Sulfaz n=10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Histo score</td>
<td>0.00 ±0.00</td>
<td>3.00 ±0.24</td>
<td>1.80 ±0.40</td>
<td>1.80 ±0.40</td>
</tr>
<tr>
<td>ICAM-1</td>
<td>0.90 ±0.30</td>
<td>4.00 ±0.34</td>
<td>2.90 ±0.30</td>
<td>2.90 ±0.30</td>
</tr>
<tr>
<td>CD62</td>
<td>0.90 ±0.30</td>
<td>3.90 ±0.30</td>
<td>1.80 ±0.40</td>
<td>1.90 ±0.30</td>
</tr>
</tbody>
</table>

### Table 4: Cytokines and oxidative stress markers immunohistochemical score

<table>
<thead>
<tr>
<th>Variable</th>
<th>Control n=10</th>
<th>Colitis n=10</th>
<th>Sild n=10</th>
<th>Sulfaz n=10</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNF-α</td>
<td>0.40 ±0.09</td>
<td>4.00 ±0.30</td>
<td>2.20 ±0.40</td>
<td>2.70 ±0.46</td>
</tr>
<tr>
<td>IL-6</td>
<td>1.00 ±0.23</td>
<td>4.00 ±0.30</td>
<td>2.10 ±0.54</td>
<td>2.50 ±0.50</td>
</tr>
<tr>
<td>MDA</td>
<td>1.70 ±0.46</td>
<td>4.00 ±0.35</td>
<td>2.30 ±0.46</td>
<td>2.70 ±0.46</td>
</tr>
<tr>
<td>MPO</td>
<td>0.80 ±0.40</td>
<td>4.00 ±0.45</td>
<td>1.80 ±0.40</td>
<td>1.70 ±0.46</td>
</tr>
</tbody>
</table>
Figure 1: Mean area of mucosal damage in control and study groups
Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; MD: Mucosal damage; Sild: Sildenafil; Sulfaz: sulfasalazine

Figure 2: Mean colonic weight (CW) in gram in control and study groups
Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine

Figure 3: Mean disease activity index (DAI) in control and study groups
Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine

Figure 4: Mean macroscopic score (MAC) in control and study groups
Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine

Figure 5: Mean histopathological score (MIC) in control and study groups
Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine

Figure 6: Histological section through colonic wall showing normal mucosal and submucosal pattern with no evidence of inflammation (arrow head) and preservation of goblet cells (arrow); A: 10X; B: 40X; H and E stain.
Figure 7: Histological section through colonic wall showing mucosal ulceration (1); superficial inflammation (2); mononuclear inflammatory infiltrate (3) and crypt abscess (4) in experimentally induced colitis in rat; A: 10X; B: 40 X; H and E stain.

Figure 8: Histological section through colonic wall showing drug effects in which there is evidence of mucosal regeneration and glandular formation, less severe inflammation and goblet cells regeneration; A: 10X; B: 40 X; H and E stain.

Figure 9: Mean intracellular adhesion molecule-1 (ICAM-1) score in control and study groups; Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild; Sildenafil; Sulfaz: sulfasalazine

Figure 10: Immunohistochemical expression of ICAM-1 showing membranous pattern (yellow arrow); A: 10X; B: 20X.

Figure 11: Mean CD62 score in control and study groups; Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild; Sildenafil; Sulfaz: sulfasalazine

Figure 12: Immunohistochemical expression of CD 62 showing membranous pattern (yellow arrow); A: 10X; B: 20X.

Figure 13: Mean TNF-α score in control and study groups; Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild; Sildenafil; Sulfaz: sulfasalazine
Figure 14: Immunohistochemical expression of TNF-α showing membranous and secretory pattern (yellow arrow); A: 10X; B: 20X.

Figure 15: Mean IL-6 score in control and study groups. Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine.

Figure 16: Immunohistochemical expression of IL-6 showing secretory pattern (yellow arrow); A: 10X; B: 20X.

Figure 17: Mean malondialdehyde (MDA) score in control and study groups. Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine.

Figure 18: Immunohistochemical expression of MDA showing cytoplasmic pattern (yellow arrow); A: 10X; B: 20X.

Figure 19: Mean myeloperoxidase (MPO) score in control and study groups. Capital letters for comparison; different letters indicates significant difference; similar letters indicates insignificant difference; Sild: Sildenafil; Sulfaz: sulfasalazine.
Effect of sildenafil on macroscopic features

Rectal instillation of acetic acid was applied in this study. The distal colon showed severe macroscopic edematous inflammation. The colonic mucosa was inflamed, hyperemic and hemorrhagic compared to normal control group. However, oral administration of sildenafil and sulfasalazine after the induction of colitis significantly (p<0.01) attenuate the colonic damage scores as shown in table 2 and figure 1. Nevertheless, sildenafil failed to produce significant reduction in the colonic weight as compared with sulfasalazine group that demonstrated significant (p<0.05) reduction effect comparable to the normal group as shown in table 2 and figure 2. However, both sildenafil and sulfasalazine showed significant decrease (p<0.01) in DAI as shown in table 2 and figure 3. Furthermore, both drugs elicit significant (P<0.01) decrease in macroscopic score as shown in table 2 and figure 4.

Effect of sildenafil on histopathological features

The present study demonstrated characteristic histological features in untreated colitis, essentially loss of intestinal crypt architecture and sloughing of intestinal cells, reduced goblet cell number and presence of different inflammatory cell infiltration as demonstrated in figure 5 and 6. On the other hand, sildenafil and sulfasalazine treated groups, the histopathological changes were significantly (P<0.05) attenuated as judged by epithelization of colonic mucosa, reduction of edema and neutrophil infiltration as shown in figure 5. No significant differences were observed between sildenafil and sulfasalazine treated groups, both of them revealed a significant decrease in the pathological scores as compared with the colitis group, figure 5 and table 3.

Effect of sildenafil on adhesion molecules (ICAM-1 and CD62)

The increased colonic ICAM-1 in the colitis group was found to be significantly (p<0.05) decreased after sildenafil and sulfasalazine treatment as shown in table (3), figure 9. Also both tested drugs cause significant (p<0.01) decrease in CD62 compared to colitis group as shown in figure 11.

Effect of sildenafil on proinflammatory cytokines (TNF-α and IL-6)

As shown in table (4) and figure 13 and 15, colonic levels of TNF-α and IL-6 showed drastic raise after acetic acid introduction compared to those of control group. In contrast these values were significantly (p<0.05) lower in rats treated with sildenafil and sulfasalazine. However, sildenafil significantly (p<0.05) decrease proinflammatory cytokines as compared with sulfasalazine treated group.

Effect of sildenafil on oxidative stress markers (MDA and MPO)

Administration of sildenafil or sulfasalazine to acetic acid treated rats significantly (p<0.05) reduced MDA compared to the colitis group as shown in figure 17. On the other hand treatment with either sildenafil or sulfasalazine significantly (p<0.01) inhibited acetic acid induced MPO production in tissue as depicted in figure 19.
and histopathological score of colon in experimentally induced colitis and this finding comparable with the finding of Khoshakhlagh et al [28] . The explanation of the beneficial role of sildenafil may be attributed to the following: first of all it has been demonstrated that inhibition of phosphodiesterase inhibitor-5 will cause marked improvement in colonic blood flow. This may be attributed to increase in the level of nitric oxide (NO) which is a potent vasodilator. The increase in NO level is attributed to accumulation of intracellular cGMP after the inhibition of the enzyme phosphodiesterase inhibitor-5 by sildenafil with resultant block to the pathway of NO degradation[30]. On the other hand, it has been shown in present study that sildenafil administration to rats experimentally induced colitis plays a significant role in reducing inflammation via reduced expression of intercellular adhesion molecules such as ICAM-1 and E-selectin. This observation has been demonstrated by Valatsou et al [31] which they proved that the immunohistochemical expression of both ICAM-1 and E-selectin were significantly reduced following administration of sildenafil. Furthermore the present study also showed significant reduction in the immunohistochemical expression of both IL-6 and TNF-α and this finding is in accordance with the findings of previous studies that has been shown that sildenafil significantly reduces the level of pro-inflammatory cytokines such as IL-6 and TNF-α [29,32]. The reduced level of pro-inflammatory cytokines may be attributed to reduced hypoxia following improvement of local blood flow caused by NO. Hypoxia is well known factor to induce expression of pro-inflammatory cytokines[33]. Moreover the present study has been shown that administration of sildenafil caused significant reduction in immunohistochemical expression of oxidative markers (MPO and MDA) and these finding agrees with previous studies that has been established that sildenafil administration significantly reduces markers of oxidative stress such as myeloperoxidase (MPO) [29,34] and MDA[29,35].

CONCLUSIONS

sildenafil has a potent anti-inflammatory and anti-oxidant effects that can be used successfully in treatment of experimentally acetic acid induced colitis in rats.

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