

# Flagellar Motility in Bacteria: Structure and Function of Flagellar Motor

Vadde Pavantheja

Saveetha School of Engineering, Saveetha Nagar, Thandalam, Chennai 602105

### Abstract:

Bacterial flagella are filamentous organelles that drive cell motion. They push cells in fluids (swimming) or on surfaces (swarming) so cells can advance toward positive situations. At the base of every flagellum, a reversible rotational engine, which is fueled by the proton-or the sodium-rationale force, is inserted in the cell envelope. The engine comprises of two sections: the turning part, or rotor, that is associated with the snare and the fiber, and the nonrotating part, or stator, that behaviors coupling particle and is liable for vitality transformation. Concentrated hereditary and biochemical investigations of the flagellum have been directed in Salmonella typhimurium and Escherichia coli, and more than 50 quality items are known to be engaged with flagellar get together and work. The vitality coupling component, be that as it may, is as yet not known. In this part, we overview our present information on the flagellar framework, in light of on considers from Salmonella, E. coli, and marine species Vibrio alginolyticus, enhanced with particular parts of other bacterial species uncovered by late investigations.

## **INTRODUCTION:**

The flagellum comprises of three sections: the fiber (helical propeller), the snare (all inclusive joint), and the basal structure (rotational engine). The biggest piece of the flagellum is the fiber, a helical structure whose shape can shift among various helical structures, a marvel named polymorphism (Asakura, 1970). This polymorphic modification of flagellar shape is related with stage variety (Iino, 1969). At the point when the cell swims, the flagellar fiber fills in as a screw propeller to change over turning movement of the engine into push (Berg and Anderson, 1973). In Salmonella, it develops to a length of around 15 mm and is made out of upwards of 30,000 duplicates of a single protein named flagellin (Minamino and Namba, 2004). A few microscopic organisms, for instance Vibrio, have a few firmly related flagellins that structure the fiber (McCarter, 2001). The flagellin subunits (FliC in Escherichia coli and Salmonella) are self-gathered to shape an empty concentric twofold rounded structure (inward and external cylinders) comprising of 11 protofilaments, which are orchestrated around parallel to the fiber pivot (Mimori et al., 1995; Morgan et al., 1995). Development of a helical structure is accomplished by a blend of the protofilaments of two particular adaptations, the R-and L-type, recognized by their helical handedness right or left (Asakura, 1970; Calladine, 1978). Every protofilament switches between these two adaptations by

reacting to an assortment of components including pH, ionic quality, mechanical pressure, and transformations (Kamiya and Asakura, 1976; Macnab and Ornston, 1977). Afterward, X-beam fiber diffraction examines uncovered somewhat extraordinary subunit pressing between the R-and L-type, whose rehash separations are 51.9 and 52.7 A°, separately (Yamashita et al., 1998).

# **TYPES OF FLAGELLA:**

There are 4 types of flagellar distribution on bacteria:

- 1. Monotrichous:
- Single polar flagellum
- Example: Vibrio cholerae

# 2.Amphitrichous:

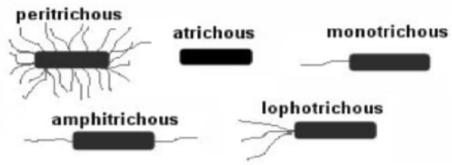
- Single flagellum on both sides
- Example: *Alkaligens faecalis*

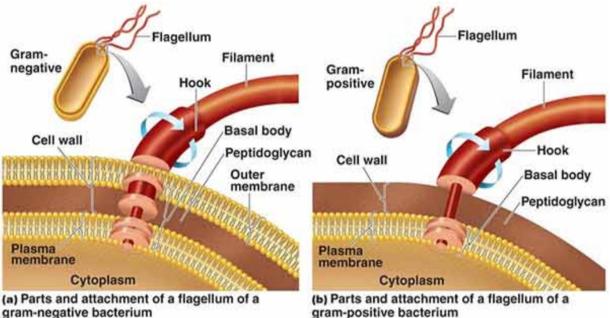
#### **3.Lophotrichous:**

- Tufts of flagella at one or both sides
- Example: Spirillum

#### 4. . Peritrichous

- Numerous falgella all over the bacterial body
- Example: Salmonella Typhi





# PARTS OF FLAGELLA:

- Each flagellum consists of three distinct parts-Filament, Hook and Basal Body.
- The **filament** lies external to the cell. •
- **Hook** is embedded in the cell envelope.

**Basal Body** is attached to the cytoplasmic membrane by ring-like structures.

# **FUNCTIONS OF FLAGELLA:**

- Movements
- Sensation
- Signal transduction
- Adhesion

Flagella are generally accepted as being important virulence factors

# **STAINING PROCESS :**

- Grow the organisms to be stained at room temperature on blood agar for 16 to 24 hours.
- Add a small drop of water to a microscope slide.
- Dip a sterile inoculating loop into sterile water
- Touch the loopful of water to the colony margin briefly (this allows motile cells to swim into the droplet of water).
- Touch the loopful of motile cells to the drop of water on the slide.
- Cover the faintly turbid drop of water on the slide with a cover slip. A proper wet mount has barely enough liquid to fill the space under a cover slip. Small air spaces around the edge are preferable.
- Examine the slide immediately under 40x for motile cells.
- If motile cells are seen, leave the slide at room temperature for 5 to 10 minutes.

- Apply 2 drops of RYU flagella stain gently on the edge of the cover slip. The stain will flow by capillary action and mix with the cell suspension.
- After 5 to 10 minutes at room temperature, examine the cells for flagella.
- Cells with flagella observed at 100x.agella may be observed

## **CONCLUSION:**

Notwithstanding the proton-driven engine, the Na+-driven engine has been examined broadly and numerous significant information have aggregated. Utilizing these bits of knowledge, a hereditary control of the Na+-driven E. coli half breed engine with fanciful stator drove us to an ongoing leap forward to watch straightforwardly the means in revolution of a solitary engine, the fundamental procedure of the engine. Starting now and into the foreseeable future, we can hope to clarify the turn instrument by examining the info and yield relations of the vitality during a solitary step in a pivot. In addition, the innovation of single-particle fluorescent perception has been presented, and it will have the option to imagine a dynamic cooperation among rotor and stator. To comprehend the component of vitality transformation that changes the particle motion into the mechanical force, the gem structures of the layer engine proteins are likewise required. We might want to take in the organic nature from the minor nanomachine of the bacterial flagella.

## **REFERENCES:**

- 1. Wang Q, Suzuki A, Mariconda S, Porwollik S, Harshey RM (June 2005). a new role of bacterial flagellum.
- 2 Bardy SL, Ng SY, Jarrell KF (February 2003). "Prokaryotic motility structures". Microbiology
- Silflow CD, Lefebvre PA (December 2001). "Assembly and 3 motility of eukaryotic cilia and flagella. Lessons from Chlamydomonas reinhardtii

- 4 Aizawa, S., Dean, G. E., Jones, C. J., Macnab, R. M., and Yamaguchi, S. (1985). Purification and characterization of the flagellar hook-basal body complex of Salmonella typhimurium. J. Bacteriol. 161, 836–849.
- 5 Akiba, T., Yoshimura, H., and Namba, K. (1991). Monolayer crystallization of flagellar L-P rings by sequential addition and depletion of lipid. Science 252, 1544–1546.
- 6 Aldridge, P., Karlinsey, J. E., Becker, E., Chevance, F. F., and Hughes, K. T. (2006). Flk prevents premature secretion of the antisigma factor FlgM into the periplasm. Mol. Microbiol. 60, 630– 643.
- Armitage, J. P. (1999). Bacterial tactic responses. Adv. Microb. Physiol. 41, 229–289.
- 8 Armitage, J. P., and Evans, M. C. W. (1985). Control of the protonmotive force in Rhodopseudomonas sphaeroides in the light and dark and its effect on the initiation of flagellar rotation. Biochimica Biophysica Acta 806, 42–45.
- Armitage, J. P., and Macnab, R. M. (1987). Unidirectional, intermittent rotation of the flagellum of Rhodobacter sphaeroides. J. Bacteriol. 169, 514–518.

- 10 Asai, Y., Kojima, S., Kato, H., Nishioka, N., Kawagishi, I., and Homma, M. (1997). Putative channel components for the fastrotating sodium-driven flagellar motor of a marine bacterium. J. Bacteriol. 179, 5104–5110.
- Asai, Y., Kawagishi, I., Sockett, R. E., and Homma, M. (1999). Hybrid motor with Hp- and Nap-driven components can rotate Vibrio polar flagella by using sodium ions. J. Bacteriol. 181, 6332– 6338.
- 12 Asai, Y., Kawagishi, I., Sockett, R. E., and Homma, M. (2000). Coupling ion specificity of chimeras between Hb- and Nab-driven motor proteins, MotB and PomB, in Vibrio polar flagella. EMBO J. 19, 3639–3648.
- B Asai, Y., Yakushi, T., Kawagishi, I., and Homma, M. (2003). Ioncoupling determinants of Nab-driven and Hb-driven flagellar motors. J. Mol. Biol. 327, 453–463.
- 44 Asakura, S. (1970). Polymerization of flagellin and polymorphism of flagella. Adv. Biophys. 1, 99–155.