

www.jpsr.pharmainfo.in

Evaluation of some biochemical and immunological parameters changes in Iraq male with Toxoplasmosis

Ahlam Jassim Taher

Department of Biology, College of Education for Pure Science (Ibn Al-Haitham), Baghdad University

Abstract

A total of 33 Iraq male positive for Toxoplasmosis and Iraq male negative for Toxoplasmosis (controls) were studies to Evaluation of some biochemical and immunological parameters changes. The parameters included lipid profile such as (Cholesterol(C), Triglycerides(TG), High-Density Lipoprotein (HDL), Low-Density Lipoprotein (LDL) and very Low -Density Lipoprotein (VLDL) and complement component C3 and C4. The results revealed significant decrease in the total cholesterol, Triglycerides, LDL and non-significant in vLDL (129.96±1.63, 130.69±2.80, 87.19±1.97, 29.24±0.83 mg/dl respectively) and non-significant increase in HDL(24.22±0.62) mg/dl compared with control group(152.07±1.63, 156.48±6.55, 99.26±1.39, 31.49±1.30 and 21.31±0.36 mg/dl). The immunological tests recorded a significant increase in C3, C4 (150.60±9.67, 31.47±1.71 mg/dl respectively) compared with control group (52.86±3.46, 15.15±0.47 mg/dl respectively). There for these results reveal that the infection with *Toxoplasma gondii* may have an essential role in alterations of lipid profile levels and complement components in infected men.

Keywords: Toxoplasma gondii, lipid profile, Toxoplasmosis, C3, C4.

INTRODUCTION

Toxoplasmosis, one of the most important common parasitic zoonosis world-wide and the obligate intracellular protozoa T. gondii is the causative agent of this disease^{1 (2)} this parasite has a complex life cycle involving sexual replication in members including both domestic and wild felids as definitive host and as exual proliferation in a wide variety of warm blooded hosts included human as intermediate host $^{(3)(4)}$. There are three infective stages of T. gondii: a rapidly dividing invasive tachyzoite, a slowly dividing bradyzoite in tissue cysts, and an environmental stage, the sporozoite, protected inside an oocyst ⁽⁴⁾ (5) (6). Human infection is mainly developed by either oral ingestion of water and foods contaminated with parasite oocysts excreted by cat feces as final host, or eating raw and undercooked meat of intermediate hosts containing tissue cysts. Moreover, the infection can be transmitted through placenta, milk, organ transplantation, and blood transfusion ^{(6) (7)}. *T. gondii* infection is widespread among humans and its prevalence varies widely from place to place approximately one-third of all humanity has been exposed to this parasite ^{(3).} Infections are usually asymptomatic in healthy individuals. But can cause severe disease in fetuses who cannot develop an effective immune response against the parasite and in immunocompromised individuals, such as AIDS patients or patients undergoing immunosuppressive therapy, can result in life-threatening disease^{(5)(8) (9)}

Parasite can enter and infect any nucleated cells, then being to growth and replicate inside a parasitophorous vacuole (PV), way out, and then infect neighboring cells. This parasites activate a potent host immune response that eliminates most of the parasite and convert back into dormant cysts witch contain bradyzoites⁽⁴⁾ (9) (10).

Lipids such as Total cholesterol, Triglycerides ,High-Density Lipoprotein, Low-Density Lipoprotein and very Low-Density Lipoprotein have been shown to play an important role in defending against parasitic infections ^(11,12) also consider important mediators of host defense during the acute phase of innate immunity. Infection and inflammation typically lower blood total cholesterol and high density lipoprotein cholesterol but increase triglycerides ⁽¹³⁾ (¹⁴⁾.

Many studies observation alteration in the levels of serum lipid during infection with intracellular parasites such as malaria ^{(15) (16}, *Leishmania* ^{(17) (18)} The complement system consists of more than 30 proteins that are either present as soluble proteins in the blood or are present as membrane-associated proteins ⁽¹⁹⁾ .Its play a major role in innate immunity where a robust and rapid response is mounted against invading pathogens. Also acting an important

role in adaptive immunity involving T and B cells that help in elimination of pathogens. $^{\rm (20)\,\,(21).}$

MATERIALS AND METHODS:

Subject's collection: The study included 60 blood samples collected from voluntaries males (33 male infected with toxoplasmosis (patients group) and 27 healthy male witch negative to toxoplasmosis as control group) who had attended to Imamein Kadhimein Medical City in Baghdad at the period March to September 2014. The ranged age between 18-52 years old. Five ml of venous blood were collected from each subject. The blood was placed in a plain tube and left to stand for 30 minutes at room temperature to clot. Then, centrifuged (3000 rpm) for 10 minutes to collect serum, which was frozen at -20°C till they were analyzed.

Biochemical Tests

Level of lipid profile included Total Cholesterol(C), Triglycerides (TGS) and High-Density Lipoprotein (HDL) were determined using a standard enzymatic assay (Linear chemicals, Montgat-Barcelona, Spain). While Low Density Lipoprotein (LDL) and very Low- Density Lipoprotein) vLDL) was calculated according to Friedewald formula ⁽¹⁷⁾

LDLmg/dl = C - HDL - TGS/5

vLDLmg/dl = TGS/5

C= Cholesterol

TGS= Triglycerides

The immunological tests

The test was carried out by using the onsite Toxo IgG / IgM (Rapid Test Kit, USA), which was a lateral flow chromatographic immunoassay for the stimultaneous detection and differentiation of IgG and IgM anti-*Toxoplasma gondii* in human sera or plasma. Complement component test is performed by using Radial immune diffusion (RID) kit (Human-Germany) for determination C3 and C4 in serum .The plate was removed from Its envelope and leaved to stand at room temperature for few minutes so that Any condensed water in the wells was evaporated. Then the wells were filled with 5µl of samples and controls and waited they have been completely Adsorbing Before handling the plate. The plate was closed and waited the required incubation period 72 hour. Measured the precipitating ring around the well after incubation and compared with conversion table that provided with the kit.

Statistical analysis:

The Data analyzed by using the software statistical packages social sciences (SPSS) version 13 and the contrast between the patients and control were analyzed by student t- test. The P \leq 0.05, P \leq 0.001 were considered to be statistically significant and results were expressed as mean \pm standard error (SE).

Table (1)	prevalence of	of anti-toxoi	olasma	antibodies	in male	volunteers
1 and (1)	prevalence	or anti-toxoj	Jiasina	antibouics	in maie	volunteers

antibodies	Nos	Positive (%)	Negative (%)
IgG	60	33 (55 %)	27(45 %)
IgM	60	0	0

Table (2) Lipid profile values of prevalence in male infected with toxoplasmosis and control

		Mean±SEM					
Group	No	Cholesterol	Triglycerides	HDL	LDL	vLDL	
		mg/dl	mg/dl	mg/dl	mg/dl	mg/dl	
patients	33	129.96± 1.63*	$130.69 \pm 2.80*$	24.22 ± 0.62	87.19± 1.97*	29.24 ± 0.83	
control	27	152.07 ± 1.63	156.48 ± 6.55	21.31±0.36	99.26± 1.39	31.49 ± 1.30	

* Significant= $P \le 0.05$

Table (3): C3 and C4 complement in male infected with toxopl	asmosis and control
--	---------------------

Group	Nos.	C3 mg/dI	C4 mg/dI
patients	33	150. $60 \pm 9.67^{**}$	31.47± 1.71**
control	27	52.86± 3.46	15.15 ± 0.47

**Significant= $P \le 0.001$

RESULTS

The Results in the Table (1)of this study show that 34 male was positive to anti-toxoplasma antibodies IgG and negative to anti-toxoplasma antibodies IgM also 26 male was negative to anti-toxoplasma antibodies IgG and IgM (control group).

DISCUSSION

The aim of this study was to assess and comparison of lipids profile and complement components in males with toxoplasmosis (patients group) and non-toxoplasmosis males (control group). Lipids are defined as organic compounds that are poorly soluble in water but miscible in organic solvents and they are play a critical role in almost all aspects of biological life they are structural components in cells and are involved in metabolic and hormonal pathways²³. The results of the Biochemical tests showed that the mean of TGS, C, LDL are significantly decrease levels and non-significant in vLDL in patients, compared with control group, While the mean of HDL seemed increased but non significantly levels in patients, compared with control group, these results were agreement with study of ^{24 25}) who revealed that in changes of lipid profile values in infected women with T. gondii. On the other hand this results were don't agree with study of Flegr which observed an increased level of cholesterol and LDL in men infected with Toxoplasmosis.

Several types of infections viral, bacterial, and parasitic have been linked to alteration in blood lipid levels Viral infection, as in human immunodeficiency virus (HIV) infections, are associated with lower blood levels of total-C and HDL-C ^{27 28} Experimental inflammation from bacterial endotoxin (lipopolysaccharide, LPS) induces similar dyslipidemias ²⁹). The hypocholesterolemia and remodeling of lipoproteins during acute phase responses of innate immunity increases clearance of LPS, particularly through increased binding of LPS to HDL particles $^{30\ 31}$.Specific parasitic infections also cause dyslipidemias. A study of the Shipibo, another indigenous Amazonian group, showed an inverse correlation of HDL-C with the density of infection by three of five parasitic worm species ³². Similar-sized samples from a city hospital in Chandigarh, India, showed lower HDL-C for patients with entamoebic and giardia parasites ³³. other studies have shown elevated levels of HDL, LDL and total cholesterol in patients suffering from parasitic infection ^{34 35} Another study were record lower in blood lipid values in patients infected with Plasmodium falciparum Malaria compared to control subjects, although the values were within the normal range also In the pretreatment, HDL , LDL and triglycerides were higher while total cholesterol was lower compared to post treatment 36 37 record abnormalities

levels of lipids characterized by decreased levels of total cholesterol, LDL, and HDL and by the increased levels of VLDL and triglycerides in children infected with *Plasmodium vivax* .Also identified Plasma lipid profile alterations like hypocholesteraemia and increased triglyceridemia are reported in patients infected with visceral leishmaniasis ^{38 18 17}.

Lipids are particular importance for pathogens, and some pathogens deliberately seek out lipid-rich host niches ³⁹⁾ or enhance the availability of lipids by manipulating the host ⁴⁰ ⁴¹. Intracellular pathogens have evolved sophisticated mechanisms to manipulate and tap into the lipid metabolism of their host cells. And Within cells of host it's often develop in specialized vacuoles and the flow of lipids between host and pathogen-controlled membranous compartments is key to the pathogen's ultimate success ⁴² ⁴³ ⁴⁴.

A numeral of previous studies have demonstrated the requirement of membrane cholesterol in host-pathogen interactions 45 46 .

Cholesterol(C) is an important component of higher eukaryotic cellular membranes and plays a crucial role in the function and the organization of membrane proteins and receptors ⁴⁷ ⁴⁸ some of which being necessary for parasite entry ⁴⁴. *Toxoplasma* cannot synthesize cholesterol novo and depends upon acquisition of LDL-derived cholesterol from the host cell, via endocytosis mediated by the LDL receptor ⁴⁹ or the LDL receptor-related protein⁵⁰. A mechanism by which host and not parasite cholesterol controls the entry of *Toxoplasma* into cells has been proposed ⁵¹. These studies indicated that cholesterol does have an important role in pathogenesis of toxoplasmosis. However, data on parasite lipid sources are scarce and the molecular mechanisms by which *Toxoplasma* acquires host cell lipids are largely indefinite ⁴⁷.

The complement system plays an essential role as a first line of host immune defense promoting the recognition, opsonization, and lysis of invading pathogens (52 ; 53) and its forms an important bridge between innate and adaptive immunity.(54).

The present results indicated that there were an increase significantly in the levels of C3 and C4 in patients compare with control .these results was agree with the results by other researches, (55 ; 56) revealed that the highest level of C3 and C4 in women with positive anti *Toxoplasma* but don't agree with Study of Al-Samarrae 57 which record highest level of C3 in patients compare with controls while lowest level of C4 in patient compare with controls but without reported significant differences. Study of Ad'hiah *et al.*, 58 record highest level of C3 and C4 in patients compare with controls while with no significant differences, Al-kalaby *et al.*, 59 record highest level of C3 in aborted women infected with *T. gondii* compare with controls with significant

differences while the highest level of C4 in patient without reported significant differences. Also Schreiber & Feldman, ⁶⁰ in vitro investigations showed that T. gondii tachyzoites are rapidly lysed by the activation of complement through the classical pathway in the presence of specific antibodies so that (Suzuki and Kobayashi, ⁶¹ prove that the presence of Ca++ is essential for the antibody-dependent cytolysis of Toxoplasma organisms, and confirm that the lytic reaction is mediated by an activation of the classical complement pathway. Hence, it is possible that the antibody-dependent killing mechanism by the activation of complement, which was observed in vitro, contributes to host defense for Toxoplasma infection in vivo by the activation of complement, which was observed in vitro, contributes to host defense for Toxoplasma infection in vivo. The collaboration between specific Antibody and presence of complement have been found capable of killing extracellular *T.gondii*⁶². All these studies indicated of an important of complement component in host defense against toxoplasma.

REFERENCES

- 1- Kucerova, P.& and Monika Cervinkova, M. (2016). Toxoplasmosis in at-risk groups of patients. Rev. Med. Microbiol., 27(1):13-19.
- 2- Wilking, H., Thamm, M., Stark, K., Aebischer, T., & Seeber, F. (2016). Prevalence, incidence estimations, and risk factors of *Toxoplasma gondii* infection in Germany: a representative, cross-sectional, serological study. Sci. Rep., 6, 22551.
- 3- Sonar, S. S. & Brahmbhatt, M.N.(2010). Toxoplasmosis: an important protozoan zoonosis. Veterinary World Vol.3(9):436-439.
- Dubey, J.P (2004).Toxoplasmosis a waterborne zoonosis. Vet.Parasitol., 126: 57–72.
- Robert-Gangneux, F., & Dardé, M.-L. (2012). Epidemiology of and diagnostic strategies for Toxoplasmosis. Clin. Microbiol.Rev., 25(2), 264–296.
- 6- Dubey, J.P.(2006). Comparative infectivity of oocysts and bradyzoites of Toxoplasma gondii for intermediate (mice) and definitive (cats) hosts. Vet. Parasitol.,140: 69–75.
- 7-Asgari, Q.; Sarnevesht, J.; Kalantari. M.; Sadat. S.J.; Motazedian, M.H.& Sarkari, B.(2011). Molecular survey of toxoplasma infection in sheep and goat from fars province, Southern Iran. Trop. Anim. Health Prod., 43:389–392.
- 8- Gov, L., Karimzadeh, A., Ueno, N., & Lodoen, M. B. (2013). Human innate immunity to *Toxoplasma gondii* is mediated by host caspase-1 and asc and parasite gra15. MBio., 4(4): e00255–13.
- 9- Carvalho, F. R., Silva, D. A. O., Cunha-Júnior, J. P., Souza, M. A., Oliveira, T. C., Béla, S. R., ... Mineo, J. R. (2008). Reverse enzymelinked immunosorbent assay using monoclonal antibodies against SAG1-related sequence, SAG2A, and p97 Antigens from *Toxoplasma gondii* to detect specific immunoglobulin G (IgG), IgM, and IgA antibodies in human sera . Clin. Vac. Immunol. : 15(8), 1265–1271.
- 10- Blader, I. J., & Saeij, J. P. (2009). Communication between *Toxoplasma gondii* and its host: impact on parasite growth, development, immune evasion, and virulence. APMIS : Acta Pathol. Microbiol. Immunol.Scand.,117(5-6): 458–476.
- Feingold, K.R.& Grunfeld, C. (2012). Lipids: a key player in the battle between the host and microorganisms. J. Lipid Res . ,53: 2560–2572 12- Vasunilashorn, S., Crimmins, E. M., Kim, J. K., Winking, J., Gurven, M., Kaplan, H., & Finch, C. E. (2010). Blood lipids, infection, and inflammatory markers in the Tsimane of Bolivia. Am. Biol. Offic. J. Hum. Biol. Coun., 22(6): 731–740.
- 13-Esteve, E.; Ricart, W.& Fernandez-Real, J.M.(2005). Dyslipidemia and inflammation: an evolutionary conserved mechanism. Clin Nutr., 24:16–31.
- 14- Jahangiri, A., de Beer, M. C., Noffsinger, V., Tannock, L. R., Ramaiah, C., Webb, N. R., ... de Beer, F. C. (2009). HDL remodeling during the acute phase response. Arterioscl. Throm. Vas. Biol., 29(2), 261–267.
- 15- Warjri, S.B.; Ete, T.; Animesh Mishra, A.; Bhupen Barman, B.; Mishra,J.; Star Pala4,S.; Taso Beyong, T. and Neel Kanth Issar, N.K. (2016). Association between Clinical Malaria and Blood Lipids in North Eastern India. Brit. J. Med. & Med. Res., 16(1): 1-7.

- 16- Visser, B. J., Wieten, R. W., Nagel, I. M., & Grobusch, M. P. (2013). Serum lipids and lipoproteins in malaria - a systematic review and meta-analysis. *Malaria Journal*, 12, 442.
- 17- Ghosh, J., Lal, C. S., Pandey, K., Das, V. N. R., Das, P., Roychoudhury, K., & Roy, S. (2011). Human visceral leishmaniasis: decrease in serum cholesterol as a function of splenic parasite load. Ann. Trop. Med. Parasitol., 105(3): 267–271.
- 18- Lal C S, Verma N., Rabidas V N, Ranjan A., Pandey K., Verma R B, Singh D., Kumar S., Das P.(2010). Total serum cholesterol determination can provide understanding of parasite burden in patients with visceral leishmaniasis infection. Clin. Chim. Acta. 411: 2112–2113.
- 19- Sarma, J.V.& Ward, P.A. (2011). The Complement System. Cell Tissue Res. 343(1): 227–235.
- 20 Kemper, C. & Atkinson, J. P. (2007). T-cell regulation: with complements from innate immunity. Nat Rev Immunol. ,7(1):9-18.
- Dunkelberger, J.R. & Song, W.C. (2010). Complement and its role in innate and adaptive immune responses. Cell Res., 20(1):34–50.
- 22- Friedewald W. T., Levy, R. I. and Fredrickson D. S. 1972. Estimation of the concentration of low-density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. Clin. Chemi., 18: 499-502.
- 23- Crook, M.A.(2012). Clinical biochemistry and metabiolic medicine 8^{th} ed . CRC Press:416 pp.
- 24-Al-Kuraishi. A.H.; Hasan,H.H.& Al-Kateeb,S.M. (2013). Lipid Profile Changes in Toxoplasmosis Aborted Women. Bagh. Sci. J., 10(1):168-175.
- 25- Flegr,J.; Príplatova,L.; Hampl,R.; Bicikoví,M.; Ripova,D.& Mohr,P.(2014). Difference of neuro- and immunomodulatory steroids and selected hormone and lipid concentrations between Toxoplasma-free and Toxoplasma-infected but not CMV-free and CMV-infected schizophrenia patients. Neuro Endocrinol Lett. 35(1): 20–27.
- 26-Al-Khamesi, M.B. (2016). Effect of Toxoplasmosis on Lipid Profile and Thyroid Hormones in Aborted Women. J. Al-Nahrain Univ.,19 (4): 122-126.
- 27-Riddler, S.A.; Li, X.; Chu, H.; Kingsley, L.A.; Dobs, A.; Evans, R.; Palella, F.; Visscher, B.; Chmiel, J.S.and Sharrett, A.R.(2007). Longitudinal changes in serum lipids among HIV-infected men on highly active antiretroviral therapy. HIV Med., 8:280–287
- 28- Rose, H.; Woolley, I.; Hoy, J.; Dart, A.; Bryant, B.; Mijch, A.and Sviridov, D.(2006). HIV infection and high-density lipoprotein: the effect of the disease vs the effect of treatment. Metabolism, 55:90– 95.
- 29- McGullicuddy, F.C.; De la Llera Moy, M.; Hinkle, C.C.; Joshi ,M.R.; Chinquoine, E.H.; Billheimer, J.T.; Rothblat, G.H.and Reilly, M.P.(2009) Inflammation impairs reverse cholesterol transport in vivo. Circulation.,119:1135–1145.
- 30- Kitchens, R.L.and Thompson, P.A.(2003). Impact of sepsis-induced changes in plasma on LPS interactions with monocytes and plasma lipoproteins: roles of soluble CD14, LBP, and acute phase lipoproteins. J. Endotox. Res., 9:113–118.
- 31- Levels, J.H.M.; Pajkrt, D.; Schultz, M.; Hoek, F.J.; Van Tol, A.; Meijers ,J.C.M. and Van Deventer ,S.J.H.(2007). Alterations in lipoprotein homeostasis during human experimental endotoxemia and clinical sepsis. Biochim Bioph. Acta, 1771:1429–1438.
- 32- Wiedermann, U.; Stemberger, H.; Unfried, E.; Widhalm, K.; Kundi, M.; Altenriederer, M.; Savedra, M. and Wiedermann, G.(1991). Intestinal worm burden and serum cholesterol or lipid concentration in a Shipibo population (Peru) Zent. Bakt., 275:279–286.
- 33- Bansal, D.; Bhatti, H. S. and Seghal, R.(2005). Altered lipid parameters in patients infected with *Entamoeba histolytica*, *Entamoeba dispar* and *Giardia lamblia*. Br. J. Biomed. Sci., 62:63– 65.
- 34- Djoumessi S. (1989). Serum lipids and lipoproteins during malaria infection. Pathol. Biol., 37: 909-11.
- 35- Faucher, J.F.; Milama, E.N.; Missinou, M.A.; Ngomo, R.; Kombila, M.and Kremsner, P.G. (2002). The impact of malaria on common lipid parameters. Parasitol. Res., 88: 1040–3.
- 36- Jacob,E.A.(2014). Assessment of altered plasma lipid pattern in *Plasmodium falciparum* malaria infected and non infected individuals. Inter. J. Hematol. Disor., 1(1): 27-30.
- 37- Dias, R.M.; Vieira, J.L.F.; Da Silva, I.R.; Feio Brasil, L.M.B.; Araújo, E.C. and Andrade, M.A. (2016). Lipid Profile of Children with Malaria by Plasmodium vivax. J. Trop. Med., 2016: 5p.

- 38- Liberopoulos, E., Alexandridis, G., Bairaktari, E., and Elisaf, M. (2002). Severe hypocholesterolemia with reduced serum lipoprotein(a) in a patient with visceral leishmaniasis. Ann. Clin. Lab. Sci. 32, 305–308.
- 39- Tanowitz, H. B., Jelicks, L. A.; Machado, F.S.; Esper, L.; Qi. X.and Desruisseaux ,M.S.(2011). Adipose tissue, diabetes and Chagas disease. Advan. Parasitol., 76:235–50.
- 40- Singh ,V.; Jamwal, S.; Jain, R.; Verma, P.; Gokhale, R. and Rao K.V.(2012). Mycobacterium tuberculosis-driven targeted recalibration of macrophage lipid homeostasis promotes the foamy phenotype. Cell host Microbe., 12:669–81.
- 41- Meester,I.; Rosas-Taraco^a, A.G.; Solís-Soto,J.M. and Salinas-Carmona, M.C. (2011). The roles of lipid droplets in human infectious disease. Med. Univer., 13(53):207–16.
- 42- Naderer, T.and McConville, M.J. (2011). Intracellular growth and pathogenesis of Leishmania parasites. Essays biochem., 51:81–95.
- Gilk, S.D.(2012). Role of lipids in *Coxiella burnetii* infection. Advan. Experim. Med. biol., 984:199–213.
- 44- Lingelbach, K.& Joiner, K.A.(1998). The parasitophorous vacuole membrane surrounding Plasmodium and Toxoplasma: an unusual compartment in infected cells. J. Cell Sci. 1998;111 (Pt 11):1467– 75.
- 45- Rosenberger, C.M.; Brumell, J.H. and Finlay, B.B. (2000). Microbial pathogenesis: lipid rafts as pathogen portals. Curr. Biol., 10(22): 823–825.
- 46- Vieira, F.S.; Corrêa, G.; Einicker-Lamas, M. and Coutinho-Silva, R. (2010) .Host-cell lipid rafts: a safe door for micro-organisms. Biol Cell 102:391–407.
- 47- Coppens ,I. (2006) Contribution of host lipids to *Toxoplasma* pathogenesis. Cell Microbiol 8:1–9.
- Paila, Y.D.and Chattopadhyay ,A .(2010). Membrane cholesterol in the function and organization of G-protein coupled receptors. Subcell Biochem., 51:439–466.
- 49- Coppens, I.; Sinai ,A. P. and Joiner, K. A. (2000). *Toxoplasma gondii* exploits host low-density lipoprotein receptor-mediated endocytosis for cholesterol acquisition. J. Cell Biol., 149: 167-180.
- 50- Portugal, L. R., ;Fernandes, L. R.; Pietra Pedroso, V. S.; Santiago, H. C.; Gazzinelli, R. T. and Alvarez- Leite, J. I. (2008). Influence of low-density lipoprotein (LDL) receptor on lipid composition, inflammation and parasitism during *Toxoplasma gondii* infection. Microbes Infect., 10: 276-284.

- 51- Coppens, I. and Joiner, K. A. (2003). Host, but not parasite cholesterol controls *Toxoplasma* cell entry by modulating organelle discharge. Mol. Biol. Cell, 14: 3804-3820.
- 52 -Lidani, K. C. F., Bavia, L., Ambrosio, A. R., & de Messias-Reason, I. J. (2017). The Complement System: A Prey of *Trypanosoma cruzi*. *Front. Microbiol.*, 8, 607.
- 53 Noris, M., & Remuzzi, G. (2013). Overview of Complement Activation and Regulation. Semin. Nephrol., 33(6):479–492.
- 54- Bennett, K.M.; Rooijakkers ,S. H.M. & Gorham, R.D. Jr. (2017). Let's tie the knot: marriage of complement and adaptive immunity in pathogen evasion, for better or worse. Front. Microbiol., 8:89.
- 55-Salloom, D. F.; AL-Warid, H.S.& Abbas, A.H. (2011). Evaluation of complements serum level C3 and C4 in pregnant women with history of toxoplasmosis. J. Biotechnol. Res. Center, 5 (2):12-16.
- 56-Shani, W. S.; Bushra, H. Sh.; Nabel, E. W. (2012). Levels of Immunoglobulins and complements in sera of patients with toxoplasmosis. Basrah J. Sci., 30(1):72-77.
- 57-Al-Samarrae, E. A. (2010). Study the Relationship of Toxoplasma IgG Titer with Some Other Immunoglobulin and Complement Components. Iraqi J. Comm. Med., 23 (2).127-129.
- 58-Ad'hiah, A. H.; Amna, N. J.; Salma, K. F. (2007). Some immunologic evaluations of toxoplasmosis in Iraqi aborted females. Um-Salama Sci. J., 4 (3):444-451.
- 59- AL- kalaby, R.F.; Sultan, B.A.& AL-Fatlawi, S.N.(2016). Assessment of C3 and C4 component of complement system in aborted women infected with *Toxoplasma gondii*. AL- Qadisiyah Med.J., 12(22):104-108.
- 60- Schreiber, R. D. & Feldman,H.A. (1980). Identification of the activator system for antibody to *Toxoplasma* as the classical complement pathway. J. Infect. Dis., 141: 366-369.
- 61- Suzuki,Y.& Kobayashi,A.(1985). Requirement for calcium ions in antibody-dependent complement-mediated cytolysis of Toxoplasma gondii. Zbl. Bakt. Hyg., 259(3): 426-431.
- 62- Hammouda, N.A.; Abo el-Naga, I.; Hussein, E.D.& Rashwan, E.A.(1995). Opsonization and intracellular killing of Toxoplasma gondii by human mononuclear phagocytes. J .Egypt Soc. Parasitol., 25(1):11-7.
- 63- Hammouda, N.A.; Abo el-Naga, I.; Hussein, E.D.& Rashwan, E.A.(1995). Opsonization and intracellular killing of Toxoplasma gondii by human mononuclear phagocytes. J .Egypt Soc. Parasitol., 25(1):11-7.