Effect Of Lisinopril (ACE Inhibitor) on Haematological and Plasma Biochemical Parameters in Male Wistar Rats

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Abstract

Objective: This study was designed to evaluate the effect of lisinopril on haematological and plasma biochemical parameters in male Wistar rats.

Methods: Twelve male rats (120 – 140 g) were divided into control (distilled water) and lisinopril-treated (0.7 mg/kg) groups (6 per group) for haematological and biochemical studies. The animals were orally treated on daily basis for 50 days. Red Blood Cell (RBC) counts and Total White Blood Cell (TWBC) counts were determined using haemocytometer. Activities of plasma Alanine Amino Transferase (ALT), Aspartate Amino Transferase (AST), Alkaline Phosphatase (ALP), as well as levels of total protein, globulin, albumin and Blood Urea Nitrogen (BUN) were determined by spectrophotometry. Data were analysed using descriptive statistics and ANOVA at p<0.05.

Results: Treatment of rats with lisinopril (0.7 mg/kg) produced significant (p<0.05) increments in PCV, Hb and RBC values relative to their respective controls.

Conclusion: It can therefore be concluded that lisinopril probably has some beneficial effects on haematological functions in male rats.

Keywords: Lisinopril, Rats, Total white blood cell counts, Red blood cell counts, Total protein.

INTRODUCTION

Lisinopril is a drug of the angiotensin-converting enzyme (ACE) inhibitor class used primarily in treatment of high blood pressure, heart failure, and after heart attacks. It is also used for preventing kidney and eye complications in people with diabetes.

Lisinopril was the third ACE inhibitor (after captopril and enalapril) and was introduced into therapy in the early 1990s [1]. A number of properties distinguish it from other ACE inhibitors: It is hydrophilic, has a long half-life and tissue penetration, and is not metabolized by the liver.

Lisinopril is typically used for the treatment of hypertension, congestive heart failure, acute myocardial infarction, and diabetic nephropathy [2].

Lisinopril has been reported to reverse the memory deficits in streptozotocin-induced experimental dementia [3]. Lisinopril significantly attenuated the oxidative damage and neuro-inflammation in the haloperidol-treated rat [4]. Lisinopril has been reported to have the potential to protect against haematological and biochemical studies. The animals were orally treated on daily basis for 50 days. Red Blood Cell (RBC) counts and Total White Blood Cell (TWBC) counts were determined using haemocytometer. Activities of plasma Alanine Amino Transferase (ALT), Aspartate Amino Transferase (AST), Alkaline Phosphatase (ALP), as well as levels of total protein, globulin, albumin and Blood Urea Nitrogen (BUN) were determined by spectrophotometry. Data were analysed using descriptive statistics and ANOVA at p<0.05.

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However, due to dearth of information from literature on the effect of lisinopril on haematological and biochemical parameters in male rats, this study therefore aims at investigating the effect of this antihypertensive agent on these aforementioned parameters in male rats.

MATERIALS AND METHODS

Experimental Animals

Adult male rats weighing between 120 g – 140 g bred in the Pre-Clinical Animal House of the College of Medicine and Health Sciences, Afe Babalola University were used. They were housed under standard laboratory conditions and had free access to feed and water; they were acclimatized for two weeks to laboratory conditions before the commencement of the experiments. All experiments were carried out in compliance with the recommendations of Afe Babalola University Ethics Committee on guiding principles on care and use of animals.

Drug

ACE inhibitors (lisinopril) tablets (TEVA UK, Ltd) were bought from Danax Pharmacy, Ibadan, Nigeria.

Lisinopril (10 mg) was dissolved in 10 ml of distilled water to give a concentration of 1.0 mg/ml.

The dosage of lisinopril used in this study was in accordance with that reported by the manufacturer.

Experimental Design

Twelve male rats (120 – 140 g) were randomly divided into two groups, with each consisting of six animals. The two groups were subjected to the following oral treatments once a day for fifty (50) days:

Group I: received 0.5 mL/100 g of distilled water as control group.

Group II: received 0.7 mg/kg of lisinopril.

Collection of blood samples

Twenty four hours (day 51) after the last dosing of all the groups, blood samples were collected from all the animals through the medial canthus with heparinized capillary tubes into EDTA bottles for hematological and plasma biochemical analysis. Before assays, the blood was centrifuged for 5 minutes using a bench top centrifuge (Centromix) and the plasma were used for the determination of the biochemical parameters.

Determination of Haematological Parameters

The red blood cells (RBC) and white blood cells (WBC) counts were determined by the Improved Neubauer haemocytometer method. The haemoglobin (Hb) concentration was determined according to [7] using the cyanomethaemoglobin method. The packed cell volume (PCV) was determined by the micro-haematocrit method according to [8]. Schilling method of differential leucocyte count was used to determine the distribution of the various white blood cells [9]. Mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were computed according to [7].

Determination of Plasma Biochemical Parameters

The total protein concentration was determined using the Biuret method [10] and the albumin concentration by the method of [11]. The globulin concentration was calculated by subtracting the albumin concentration from the total protein concentration. Activities of plasma alanine transaminase (ALT) and aspartate...
transaminase (AST) were determined according to the method of [12]. The level of creatinine, urea and alkaline phosphatase were determined using the method of [13]. All the above biochemical parameters were determined in the plasma using the Randox kits.

**Statistical Analysis**

The mean and standard error of mean (S.E.M.) were calculated for all values. Comparison between the control and experimental groups was done using one-way analysis of variance (ANOVA) with Duncan’s Multiple Range Test. Differences were considered statistically significant at p<0.05.

**RESULTS**

The effect of lisinopril (0.7 mg/kg) on haematological and plasma biochemical parameters after treatment of rats for 50 days is shown in Tables 1 and 2 respectively.

Treatment of rats with lisinopril (0.7 mg/kg) caused significant (p<0.05) increments in PCV, Hb and RBC values relative to their respective controls.

Treatment of rats with lisinopril (0.7 mg/kg) produced no significant (p>0.05) changes in TWBC, platelet, lymphocyte, neutrophil, monocyte, eosinophil, MCV, MCHC and MCH values relative to their respective controls.

**DISCUSSION**

The result of the haematological study has shown that lisinopril caused significant increments in the PCV, Hb and RBC values. This could indicate that the drug has the potential to stimulate erythropoietin release from the kidney which is the humoral regulator of RBC production [14]. It could also indicate that there were increments in the oxygen carrying capacity of the blood and the amount of oxygen delivered to the tissues, since RBC and haemoglobin (Hb) are very important in transferring respiratory gases [15]. It has been reported that values of RBC and associated parameters lower than normal ranges are indicative of anaemia conditions while higher values are suggestive of polycythemia [16]; thus, the drug may have the potential to induce polycythemia. Also, the drug may have adverse effects on the bone marrow, kidney and haemoglobin metabolism, since it has been reported that only substances which significantly affect the values of red blood cells and associated parameters would have effects on the bone marrow, kidney and haemoglobin metabolism [17].

Lisinopril caused no significant change in TWBC value which probably indicates that it has no effect on the ability of the body to defend against invading organisms [18]. A similar result was reported by [19] in *Viscum album* extract treated rats.

Lisinopril caused no significant change in the platelet values which probably indicates that it has no effect on the haemostatic function of the body. Contrary result was reported by [20] in *Fadogia agrestis* extract treated rats.

Lisinopril caused no significant change in lymphocyte value which probably indicates that it has no effect on the acquired immune response of the body. Similar result was reported by [21] in isolated ergosterol treated rats.

Lisinopril caused no significant increase in the neutrophil count which probably indicates it has no effect on the ability of the body to attack and destroy invading bacteria, viruses and other injurious agents (phagocytosis). Contrary result was reported [22] in *Dennettia tripetala* extract treated rats. Lisinopril caused no significant change in the monocyte value which probably indicates that it has no effect on the phagocytic function of the body [23]. Contrary result was reported by [24] in *Saccharomyces cerevisiae* extract fed hens.

Lisinopril caused no significant change in eosinophil value which could indicate that it has no effect on the anti-allergic and anti-parasitic infectious responses of the body. Contrary result was reported by [25] in *Arctotis acutostides* extract treated rats and mice.

Lisinopril caused insignificant changes in the MCV and MCH values which probably indicates that it has no effect on induction of macrocytic anaemia, since increased MCV and MCH values are known to be indicative of macrocytic anaemia [26]. Similar result was reported by [27] in *Jatropha gossypifolia* extract treated rats. The insignificant change in the MCHC value caused by lisinopril probably indicates that it has no effect on induction of hereditary spherocytosis, since MCHC values are known to be elevated in hereditary spherocytosis. Similar result was reported by [27] in *Jatropha gossypifolia* extract treated rats. The insignificant change in the MCHC value caused by lisinopril probably indicates that it has no effect on induction of hereditary spherocytosis, since MCHC values are known to be elevated in hereditary spherocytosis. Similar result was reported by [27] in *Jatropha gossypifolia* extract treated rats. The insignificant change in the MCHC value caused by lisinopril probably indicates that it has no effect on induction of hereditary spherocytosis, since MCHC values are known to be elevated in hereditary spherocytosis. Similar result was reported by [27] in *Jatropha gossypifolia* extract treated rats.

The result of the plasma biochemical study has shown that treatment of rats with lisinopril caused insignificant changes in total protein level. This might indicate that the drug has no effect on the buffering capacity of the blood as well as having no effect on colloid osmotic pressure, since plasma proteins have been reported to be responsible for 15% of buffering capacity of blood [18] and that osmotic pressure caused by the plasma proteins (called colloid osmotic pressure) tends to cause fluid movement by osmosis. Contrary result was reported by [28] in *Euphorbia heterophylla* extract treated rats.

Lisinopril caused no significant changes in albumin level which probably indicates that it has no effect in the plasma levels of metals, ions, fatty acids, amino acids, bilirubin and enzymes.

**Table 1: Effect of 50 days treatment with lisinopril on haematological parameters in male rats**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>Lisinopril (0.7 mg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCV (%)</td>
<td>43.40 ± 0.52</td>
<td>46.33 ± 0.63*</td>
</tr>
<tr>
<td>Hb (g/dL)</td>
<td>14.38 ± 0.34</td>
<td>15.23 ± 0.21*</td>
</tr>
<tr>
<td>RBC (&lt;10⁶/µL)</td>
<td>7.31 ± 0.32</td>
<td>7.64 ± 0.15*</td>
</tr>
<tr>
<td>TWBC (&lt;10⁶/µL)</td>
<td>3.70 ± 0.21</td>
<td>4.65 ± 0.41</td>
</tr>
<tr>
<td>Platelets (&lt;10⁵/µL)</td>
<td>1.39 ± 0.05</td>
<td>1.10 ± 0.03</td>
</tr>
<tr>
<td>Lymphocytes (%)</td>
<td>71.00 ± 0.83</td>
<td>69.17 ± 0.82</td>
</tr>
<tr>
<td>Neutrophils (%)</td>
<td>25.40 ± 0.75</td>
<td>27.17 ± 0.76</td>
</tr>
<tr>
<td>Monocytes (%)</td>
<td>1.60 ±0.11</td>
<td>2.00 ±0.16</td>
</tr>
<tr>
<td>Eosinophils (%)</td>
<td>2.00 ±0.16</td>
<td>1.67 ± 0.14</td>
</tr>
<tr>
<td>MCV (FL)</td>
<td>59.36 ± 0.55</td>
<td>60.66 ± 0.41</td>
</tr>
<tr>
<td>MCHC (g/dL)</td>
<td>33.12 ± 0.41</td>
<td>32.90 ± 0.20</td>
</tr>
<tr>
<td>MCH (pg)</td>
<td>19.67 ± 0.34</td>
<td>19.95 ± 0.21</td>
</tr>
</tbody>
</table>

(n=6, *p<0.05)

**Table 2: Effect of 50 days treatment with lisinopril on plasma biochemical parameters in male rats**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>Lisinopril (0.7 mg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Protein (g %)</td>
<td>6.80 ± 0.13</td>
<td>7.20 ± 0.16</td>
</tr>
<tr>
<td>Albumin (gm %)</td>
<td>2.66 ± 0.13</td>
<td>2.67 ± 0.18</td>
</tr>
<tr>
<td>Globulin (gm %)</td>
<td>4.14 ±0.18</td>
<td>4.53 ± 0.24</td>
</tr>
<tr>
<td>AST (µ/L)</td>
<td>42.20 ± 0.87</td>
<td>40.67 ± 0.91</td>
</tr>
<tr>
<td>ALT (µ/L)</td>
<td>29.60 ± 0.74</td>
<td>30.33 ± 0.78</td>
</tr>
<tr>
<td>ALP (IU/L)</td>
<td>110.00 ± 1.71</td>
<td>111.00 ± 1.47</td>
</tr>
<tr>
<td>BUN (mg/dL)</td>
<td>15.56 ± 0.28</td>
<td>16.35 ± 0.31</td>
</tr>
<tr>
<td>Creatinine (µmol/L)</td>
<td>0.74 ± 0.02</td>
<td>0.72 ± 0.03</td>
</tr>
</tbody>
</table>

(n=6, *p<0.05)
since it has been reported that albumin serves as a carrier for drugs [18]. Contrary result was reported by [29] in Jatropha gossypifolia extracts treated rats.

Lisinopril produced no significant changes in globulin level which probably indicates that it has no effect on both the natural and acquired immunity of the body against invading organisms, since it has been reported that globulins are principally responsible for the body’s both natural and acquired immunity against invading organisms [23]. Similar result was reported by [30] in Portulaca oleracea extracts treated rats.

The insignificant change in the activity of AST caused by the drug could indicate that it has no effect on induction of tissue necrosis, since it has been reported that elevation in the activity of AST can indicate that it has no effect on induction of tissue necrosis, because it has been reported that albumin serves as a carrier for albumin [18]. Co ntrary result was reported by [29] in Jatropha gossypifolia extracts treated rats.

Lisinopril caused no significant change in ALP level. This probably indicates the absence of cholestasis, since ALP has been reported that ALP is present in the liver and other cells and is particularly useful in measuring hepatic necrosis, especially in small animals [33]. Contrary result was reported by [34] in Moringa oleifera extract treated rats.

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CONCLUSION

In conclusion, this study has shown that lisinopril has some beneficial effects on the haematological functions in male rats. However, the effect of this antihypertensive agent on human haematological function and blood chemistry are unknown; nevertheless, considering these findings in animal model, it is recommended that patients should strictly comply with the dosage regimen as recommended by their physicians.

CONFLICT OF INTEREST

We vehemently declare that there is no conflict of interests in this research work.

REFERENCES


